Lab Dept:

Microbiology

Test Name: OVA AND PARASITE EXAM, ASPIRATED SPECIMEN

General Information

Lab Order Codes:	OAP
Synonyms:	Duodenal aspirate for Ova and Parasites; O & P Exam, duodenal aspirate; O & P Exam, sputum; O & P Exam, urine; Schistosoma Exam, urine
CPT Codes:	87177 – Ova and parasites, direct smears, concentration and identification 88312 – Special stains; Group I for microorganisms, each
Test Includes:	Examination of aspirate for intestinal parasites by direct/concentrated microscopic exam and trichrome stain.
	If only Cryptosporidium or Giardia lamblia are requested, refer to specific listing for Cryptosporidium/Giardia FA.
Logistics	
Lab Testing Sections:	Microbiology
Phone Numbers:	MIN Lab: 612-813-5866
	STP Lab: 651-220-6555
Test Availability:	Daily, test performed 0700 – 1500
Turnaround Time:	1 - 2 days
Special Instructions:	 Specimen site and date/time of collection are required for specimen processing. Indicate travel history, clinical diagnosis or parasite suspected on requisition.
	Warning: Aspirated specimens collected from a patient infected with parasites are highly infectious. Use extreme caution when <i>Entamoeba histolytica</i> , <i>Hymenolepsis nana</i> , and <i>Taenia</i> sp. are suspected.
Specimen	

Specimen Type:

Aspirate of cyst or abscess, duodenal aspirate, mid-day urine, sigmoid aspirate, or early morning sputum

Container:	Sterile, leak-proof container
Volume:	3 - 4 mL aspirate, 1-3 mL sputum or entire mid-day urine collection with last voided portion containing blood and mucus.
Collection:	Duodenal Aspirate:
	 Onsite collections ONLY: 1. Specimen is obtained by use of a gastroduodenal tube or a fiber optic endoscopy study, either by direct aspiration or into a trap. 2. Place aspirate into a sterile leak-proof container. 3. Transport directly to the laboratory (≤15 minutes) since specimens must be examined within 1 hour of collection.
	Sigmoidoscopy:
	 Onsite collections ONLY: 1. Perform flexible or rigid sigmoidoscopy. 2. Aspirate liquid from the inflamed bowel with a pipette passed through the sigmoidoscope. 3. Place aspirate into a leak-proof container. 4. Transport directly to the laboratory (≤15 min) since specimens must be examined within 1 hour of collection.
	Sputum (Expectorate):
	 Onsite collections ONLY: 1. Collect early morning specimen under the direct supervision of a nurse or a physician. 2. Have patient rinse or gargle with water to remove superficial flora. 3. Instruct patient to cough deeply to produce a lower respiratory specimen. 4. Exam specimen to make sure it contains thick mucus. Do not submit saliva. 5. Transport directly to the laboratory (≤15 min) since specimens must be examined within 1 hour of collection.
	Urine:
	 Onsite collections ONLY: 1. Collect mid-day urine specimen in a sterile container. Peak egg excretion occurs between noon and 3 p.m. 2. For patients with hematuria, eggs are associated with the terminal (last voided) portion of the specimen containing blood and mucus. 3. Transport directly to the laboratory (≤15 min) since specimens must be examined within 1 hour of collection.
Transport/Storage:	Transport to the Microbiology Laboratory immediately at room temperature. Do not refrigerate specimen. Refrigeration inhibits the motility of certain parasites.

Sample Rejection:	Specimen with a transit time exceeding 1 hour after collection; specimen not submitted in appropriate transport container; improperly labeled specimen; insufficient volume; external contamination. If an unacceptable specimen is received, the physician or nursing station will be notified and another specimen will be requested before the specimen is discarded.
Interpretive	
Reference Range:	No ova or parasites seen.
Limitations:	This procedure does not detect <i>Cryptosporidium parvum</i> . Refer to the specific listing for <u>Cryptosporidium/Giardia FA</u> .
Methodology:	Concentrated microscopic exam and trichrome stain
References:	Cook, JH, and M Pezzlo (1992). Specimen receipt and accessioning. Section 1. Aerobic bacteriology, 1.2.1-4. In HD Isenberg (ed) Clinical Microbiology Procedures Handbook. American Society for Microbiology, Washington DC
	Miller, J Michael (1999) A Guide To Specimen Management in Clinical Microbiology, American Society for Microbiology, Washington DC
	Miller, J Michael, and HT Holmes (1999) Specimen Collection, Transport, and Storage In PR Murray et al, (ed), Manual of Clinical Microbiology, 7 th edition, American Society for Microbiology, Washington DC, pp 33-104
Updates:	11/20/2014: Onsite collections ONLY