Lab Dept: Coagulation

Test Name: VON WILLEBRAND SCREEN

**General Information**

Lab Order Codes: VWS

Synonyms: See individual test listings, links can be found under Reference Range

CPT Codes:
- 85240 - Clotting; factor VIII (AHG), one stage
- 85245 - Factor VIII vW Factor, Ristocetin cofactor
- 85246 – Factor VIII vW antigen factor

Test Includes:
Factor VIII, Ristocetin Cofactor and von Willebrand antigen reported as a % and Ristocetin Cofactor:vW Antigen Ratio.

A specimen will be held for possible Multimer analysis, please contact Pathology at 612-813-6711 for consultation. Tubes will be held for one month.

**Logistics**

Test Indications: Useful for diagnosis (or exclusion) of von Willebrand’s disease or trait.

Lab Testing Section: Coagulation (Minneapolis Campus)

Phone Numbers:
- MIN Lab: 612-813-6280
- STP Lab: 651-220-6550

Test Availability: Daily, 24 hours

Turnaround Time: 3 – 5 Days, performed Tuesday and Thursday only

Special Instructions: See Patient Preparation

Deliver immediately to the laboratory at room temperature. Indicate when specimen is drawn from a line or a heparin lock.

**Specimen**

Specimen Type: Whole blood

Container: Light Blue (Buffered Na Citrate 3.2%) top tubes

Draw Volume: Minimum: 5.4 mL blood (into two 2.7 mL tubes)
Processed Volume: Minimum: 1.8 mL plasma

Collection:
- A clean venipuncture is essential, avoid foaming.
- Entire sample must be collected with single collection, pooling of sample is unacceptable.
- Capillary collection is unacceptable.
- Patient’s with a hematocrit level >55% must have a special tube made to adjust for the hematocrit; contact lab for a special tube.
- Mix thoroughly by gentle inversion. Deliver immediately to the laboratory at room temperature via courier or pneumatic tube.

Off campus collections:
- Must be tested within 4 hours.
- Do not refrigerate.
- If not received in our lab within 4 hours of collection, sample must be centrifuged and *platelet-poor plasma removed from cells and transferred to an aliquot tube being careful not to disturb the cell layer. Centrifuge the plasma a second time and transfer into a clean aliquot tube being careful not to include any residual platelets on the bottom of the tube. Freeze at -20°C and deliver to the lab on dry ice within 2 weeks.
*Validation of your lab's centrifuge for platelet poor plasma is required

Special Processing: Lab Staff: All testing will be performed in Minneapolis.

St. Paul Lab: Send whole blood specimens to Mpls for processing via courier. Must be processed within 4 hours of collection. Contact Mpls prior to sending.

Mpls Lab: Centrifuge specimen, remove plasma sample taking care not to disturb buffy coat. Perform a second spin after removing plasma from the original collection containers. Aliquot into two screw top plastic vials (Minimum: 0.5 mL in each). Store frozen at -70 degrees C (one vial in “VWS to be completed” cup and the other in the “von Willebrand Multimer Testing” cup for potential Multimer analysis) temperatures.

Patient Preparation: No Heparin for 2 days prior to testing.

Sample Rejection: Improper tube; clotted sample; under-filled tube; mislabeled or unlabeled specimens

Interpretive

Reference Range: See individual tests for specific reference ranges:

von Willebrand Antigen
Factor VIII Assay
Ristocetin Cofactor

For further interpretation of results or questions regarding Multimer analysis, please contact Pathology at 612- 813- 6711.
<table>
<thead>
<tr>
<th><strong>Critical Values:</strong></th>
<th>N/A</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Limitations:</strong></td>
<td>See individual test listings, links can be found under Reference Range</td>
</tr>
<tr>
<td><strong>Methodology:</strong></td>
<td>See individual test listings, links can be found under Reference Range</td>
</tr>
<tr>
<td><strong>References:</strong></td>
<td>See individual test listings, links can be found under Reference Range</td>
</tr>
</tbody>
</table>
| **Updates:**        | 5/24/2010: Tubing patient specimens is no longer prohibited.  
                       9/15/2014: Added Off Campus collection info.  
                       8/2/2015: Days performed updated  
                       12/2/2019: Updated draw volume. |