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**Lab Dept:** Microbiology/Virology

**Test Name:** CHLAMYDIA PNEUMONIAE DNA PCR

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***General Information***

**Lab Order Codes:** CHLP

**Synonyms:** Culture, Chlamydia pneumoniae (TWAR); Culture, TWAR (Chlamydia pneumoniae); PCR, Chlamydia pneumoniae (TWAR)

**CPT Codes:** 87486 - Infectious agent detection by nucleic acid (DNA or RNA); Chlamydia pneumoniae, amplified probe technique

**Test Includes:** Polymerase Chain Reaction (PCR). Culture for *Chlamydia pneumoniae* has been replaced by DNA PCR.

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***Logistics***

**Lab Testing Sections:** Microbiology - Sendouts

**Referred to:** LabCorp/Viomed (LC Test# 138263)

**Phone Numbers:** MIN Lab: 612-813-5866

STP Lab: 651-220-6555

**Test Availability:** Daily, 24 hours, Set up on Monday and Thursday

**Turnaround Time:** 3 – 5 days

**Special Instructions:** Requisition must state **specific site** of specimen and **date/time of collection**.

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***Specimen***

**Specimen Type:** Swab: Throat or nasopharyngeal swab in M4 VTM transport media

Nasal wash or aspirate: Place in sterile container.

Bronchial washings or bronchoalveolar lavage (1 mL) in a sterile container

**Container:** Container dependent upon specimen collected:

**Onsite collections:** Swab or sterile container

Offsite collections: M4 VTM transport media (available from Microbiology) or sterile container

**Volume:**

**Bronchoscopy specimens:** 1 mL

**Nasopharyngeal swabs:** 2 minitip swabs; one each from the right and left nares

**Nasal Washings/Nasal aspirate:** 1 mL

**Throat swab:** 1 swab

**Collection:****Bronchoscopy:**

1. Specimen obtained by physician through the biopsy channel of the bronchoscope.
2. Transfer specimen into a sterile container.

**Nasopharyngeal Swabs:**

1. Carefully insert a minitip swab into the posterior nasopharynx via the nose, collecting one each from right and left nares.
2. Rotate the swab slowly for 5 s to absorb secretions, keeping the swab near the floor and the septum of the nose.

**Throat Swabs:**

1. Depress tongue with a tongue depressor.
2. Sample the posterior pharynx, tonsils, and inflamed areas with a sterile swab.

**Nasopharyngeal Washings:**

1. Tilt patient's head back at a 70° angle.
2. Insert rubber bulb syringe containing 1 – 2 mL of sterile saline until it occludes the nostril.
3. Collect specimen (Minimum: 1 mL) with one complete squeeze and release bulb.
4. Repeat in other nostril.
5. Place washings in container.

**Nasal Aspiration:**

1. Prepare suction set up on low to medium suction.
2. Wash hands and put on protective barriers (e.g., gloves, gown, mask).
3. Place child supine and obtain assistant to hold child during

procedure.

4. Attach luki tube to suction tubing and #6 French suction catheter.

5. Insert catheter into nostril and pharynx without applying suction.

6. Apply suction as catheter is withdrawn.

If necessary, suction 0.5 - 1 mL of normal saline through catheter in order to clear the catheter and increase the amount of specimen in the luki tube.

7. Place aspirate in sterile container.

**Special Processing:**

**Offsite collections:**

Throat swab: Place in M4 VTM, break off plastic shaft. Allow the swab to remain in the tube.

2 NP swabs: Place both swabs in M4 VTM, cut the wire shafts of the swabs. Allow the swabs to remain in the tube.

**Transport/Storage:**

**Onsite collections:** Transport to the Microbiology Laboratory immediately.

**Offsite collections:** Refrigerate specimen.

Specimens must be promptly transported to the laboratory, with the next available courier, not to exceed 24 hours from the time of the collection. However, delayed transport causes a delay of test results.

**Sample Rejection:**

Improperly labeled specimen; specimens with prolonged transit time (see [Transport/Storage](#) for requirements); specimen not submitted in appropriate transport container; insufficient volume; external contamination. If an unacceptable specimen is received, the physician or nursing station will be notified and another specimen will be requested before the specimen is discarded.

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**Interpretive**

**Reference Range:**

No *Chlamydia pneumoniae* detected by DNA PCR

**Limitations:**

A negative result does not rule out the presence of PCR inhibitors in the patient specimen or *C. pneumoniae* nucleic acid concentrations below the level of detection of the assay.

**Methodology:**

DNA PCR

**References:**

LabCorp Web Page [www.labcorp.com](http://www.labcorp.com) or <http://www.viomed.com/>  
November 2014

**Updates:**

3/6/2009: Reference laboratory will no longer accept tissue samples for this testing. Added info for nasopharyngeal washings and nasal aspirations.

8/21/2013: Update to M4 VTM, removal of universal media.

11/11/2014: Addition of offsite collection information.